This document is the Accepted Manuscript version of a Published Work that appeared in final form in Environmental Science & Technology, copyright © American Chemical Society after peer review and technical editing by the publisher. To access the final edited and published work see http://dx.doi.org/10.1021/acs.est.0c03113





Subscriber access provided by NIBIO (Norwegian institute of Bioeconomy Research)

Ecotoxicology and Public Health

Wastewater-aged silver nanoparticles in single and combined exposures with titanium dioxide affect the early development of the marine copepod *Tisbe battagliai*

Anastasia Georgantzopoulou, Julia Farkas, kuria ndungu, Claire Coutris, Patricia Almeida Carvalho, Andy M. Booth, and Ailbhe Macken

Environ. Sci. Technol., Just Accepted Manuscript • DOI: 10.1021/acs.est.0c03113 • Publication Date (Web): 27 Aug 2020

Downloaded from pubs.acs.org on August 28, 2020

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.

is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

1	Wastewater-aged silver nanoparticles in single and
2	combined exposures with titanium dioxide affect
3	the early development of the marine copepod
4	Tisbe battagliai
5	Anastasia Georgantzopoulou, *ª Julia Farkas, ^b Kuria Ndungu, ^a Claire Coutris, ^d Patricia
6	Almeida Carvalho, ^c Andy M. Booth ^b and Ailbhe Macken ^a
7	^a NIVA, Norwegian Institute for Water Research, Gaustadalleen 21, 0349, Oslo, Norway
8	^b SINTEF Ocean, Brattørkaia 17C, 7010, Trondheim, Norway
9	^c SINTEF Materials and Chemistry, Forskningsveien 1, 0373, Oslo, Norway
10	^d NIBIO Norwegian Institute of Bioeconomy Research, Høyskoleveien 7, 1431 Ås, Norway
11	
12	
13	

14 ABSTRACT

15 In this study, the effects of aged Ag and TiO₂ NPs, individually and as a mixture, in wastewater relative to their pristine counterparts on the development of the copepod nauplii (Tisbe 16 17 battagliai) were investigated. NP behavior in synthetic wastewater and seawater was characterized during aging and exposure. A delayed development and subsequent mortality 18 was observed after 6 days of exposure to aged Ag NPs, with a 2-fold decrease in EC50 (316 19 20 $\mu g/L$) compared to pristine NPs (EC50 640 $\mu g/L$) despite the similar dissolved Ag concentrations measured for aged and pristine Ag NPs (441 µg/L and 378 µg/L, respectively). 21 In co-exposures with TiO₂ NPs, higher dissolved Ag levels were measured for aged NPs (238.3 22 23 μ g/L) relative to pristine NPs (98.57 μ g/L). Co-exposure resulted in a slight decrease (15%) in the Ag NP EC50 (270 µg/L) with a 1.9-fold increase in Ag NP retained within the organisms 24 after depuration (2.82% retention) compared to Ag NP single exposures as measured with sp-25 ICP-MS suggesting that the particles are still bioavailable despite the heteroaggregation 26 observed between Ag, Ti NPs and wastewater components. This study shows that the presence 27 28 of TiO₂ NPs can affect the stability and toxicity of Ag NPs in complex media that cannot be predicted solely based on ionic, total or nanoparticulate concentrations and the need for 29 studying NP interactions in more complex matrices is highlighted. 30

31

33 INTRODUCTION

Silver (Ag) and titanium dioxide (TiO₂) nanoparticles (NPs) are used in numerous industrial 34 35 applications and the number of consumer products incorporating Ag and TiO₂ NPs has increased over recent years^{1,2}. Both Ag and TiO₂ NPs are released into wastewater streams 36 during the life cycle of such products, reaching wastewater treatment plants (WWTPs). As a 37 result, NPs in consumer products are likely to be released into the aquatic environment through 38 effluent discharges or through direct release into surface waters during product use (e.g. 39 sunscreens). Several studies have shown the presence of nanosized Ag and TiO₂ particles in 40 WWTPs^{3,4}, effluents and surface waters⁵. 41

In different environmental matrices, NPs can undergo a number of environmental 42 43 transformations (aging), including changes in agglomeration/aggregation, oxidation, dissolution, precipitation of other forms, surface modification and interaction with (in)organic 44 species/ligands^{6,7}. Aging and transformation of NPs in environmental media typically leads to 45 NPs with altered physicochemical properties that differ from their pristine forms. This can 46 result in subsequent changes in particle behavior, leading to differences in fate, uptake and 47 48 subsequent effects that are difficult to predict from information on the pristine particles alone. Laboratory exposures with well-dispersed pristine NPs are therefore not expected to represent 49 realistic environmental conditions for effects assessment. Several studies have stressed the 50 51 need to assess the effects of NPs in more relevant and complex matrices such as wastewaters and effluents. It has also been suggested that aging of NPs prior to ecotoxicological assessment 52 can minimize further transformation during exposures and allow particle behavior to be studied 53 54 under more environmentally realistic conditions⁸. Despite this, the majority of ecotoxicological studies are still conducted in simplified/standardized media, with only a limited number of 55 exposure studies incorporating transformed materials being reported^{7,9–13}. 56

Several studies have described the specific changes or transformation that may occur to specific 58 59 NP materials in various environmental matrices. The chemical transformation of Ag NPs to nanoscale Ag₂S species has been reported in WWTPs¹⁴. In Cl⁻-rich media, oxidative dissolution 60 is followed by the formation of different AgCl species, which have been shown to remain 61 bioavailable to aquatic organisms¹⁵. A study using CuO NPs showed that transformation within 62 a laboratory septic tank led to decreased toxicity in the zebrafish embryo hatching assay due to 63 formation of non-bioavailable Cu species¹⁰ while a decreased bioavailability and toxicity of 64 wastewater-borne Ag NPs was previously shown for rainbow trout¹⁶. In contrast, effluents from 65 Ag NP-spiked lab-scale sewage treatment plant exhibited higher toxicity to zebrafish embryos 66 than the pristine NPs¹¹. Another study using a lab-scale WWTP spiked with Ag and TiO₂ NPs 67 (co-exposure) for 5 weeks has shown that Ag particles were still present in the effluent and 68 69 despite the observed Ag NP transformation, they were found to impact the survival of the marine copepod Tisbe battagliai and led to increased oxidative stress levels, increased 70 paracellular permeability and altered defense responses to the RTgill-W1 cell line model⁹. It 71 has recently been shown that Ag NPs were still bioavailable to soil microorganisms and 72 terrestrial isopods, despite undergoing sulfidation^{17,18}. 73

It has been highlighted that the behavior and fate of NPs in environmental media depends on the size/aggregate size, concentration and media composition^{8,19–21}. Therefore, in depth characterization of the exposure conditions is essential for an improved understanding of particle interactions with organisms and for assessing the environmental hazard. Moreover, different NP types will most likely co-exist in WWTPs and receiving environmental compartments and their interaction can result in further transformation and altered bioavailability and toxicity, yet studies investigating the combined effects of multiple NPs are
very scarce^{9,22,23}.

82 The aim of the current study was to elucidate differences in the toxicity of pristine and wastewater-aged Ag NPs and TiO₂ NPs and their mixture using sensitive life stages of the 83 epibenthic marine copepod T. battagliai. A pre-aging step in synthetic wastewater (4h) was 84 incorporated to account for rapid transformations previously demonstrated for NPs in 85 wastewater. Pristine Ag NPs and TiO₂ NPs were used 'as received' for comparison. The NP 86 behavior and characteristics during the aging process and during the exposure period was 87 studied using a combination of imaging and analytical techniques, including dynamic light 88 scattering (DLS) and Laser Doppler electrophoresis (size and agglomeration/aggregation, 89 surface charge), Ultraviolet-visible spectroscopy (UV-Vis, stability in different media) and 90 inductively coupled plasma mass spectrometry (ICP-MS, Ag dissolution and concentration 91 determination). Single particle (sp)-ICP-MS combined with an enzymatic digestion method 92 was used to determine the NP concentration retained in the organisms at the end of the exposure 93 94 period. The effects of aged vs pristine NPs on the early life stage development of T. battagliai was studied over 6 days of exposure. T. battagliai was chosen as a model organism due to their 95 96 importance and ecological significance in food webs, short life cycle, distinctive developmental stages, relatively high sensitivity and the relevance of epibenthic organisms to NP exposure⁹. 97

98 MATERIALS AND METHODS

99 Nanoparticle preparation.

Polyvinylpyrrolidone (PVP) coated Ag NPs (Econix 25 nm, Nanocomposix, Czech Republic)
 were supplied as aqueous dispersion (no other dispersant was present), while TiO₂ NPs (NM 101, 5 nm primary particles, Joint Research Centre Repository, Ispra, Italy) were supplied in
 powder form and dispersions prepared as previously described⁹ and according to the FP7 EU

NANoREG sonication protocol²⁴. Briefly, TiO₂ NP stocks (2.56 mg/ml in 0.22 µm filtered 104 MilliQ water) were prepared by sonicating in an ice water bath for 13 min using a calibrated 105 probe sonicator. The NP stocks were characterized with scanning transmission electron 106 microscopy (STEM), single particle ICP-MS (sp-ICP-MS) and dynamic light scattering (DLS, 107 Zetasizer nano ZS Malvern Instruments, UK) as described previously⁹. To prepare the exposure 108 dispersions, the stock dispersions were either diluted directly in filtered (0.22 µm) natural 109 110 seawater (referred to as pristine NPs), or were aged in synthetic wastewater (SWW) before being diluted in seawater (referred to as aged NPs). The concentrations used for the aging 111 112 process were 10 mg/L for Ag NPs, 100 mg/L for TiO₂ NPs and these were combined for the NP mixture (10 and 100 mg/L Ag and TiO2 NPs, respectively). The Ag NP and TiO2 NP 113 concentrations used in the current study are higher than the levels currently being measured in 114 WWTP influents and surface waters. The justification for the selected concentrations was the 115 limitations with DLS characterization in complex matrices. In complex matrices such as 116 wastewater or SWW there is interference caused by the high particle background concentration. 117 A ten times higher concentration of TiO₂ NPs relative to Ag NPs was used due to the higher 118 concentrations of TiO₂ NPs commonly found in environmental samples and WWTPs. 119

120 NP aging in synthetic wastewater and characterization.

Exposure dispersion preparation. To represent the rapid transformation of NPs that occurs in wastewater, Ag NPs, TiO₂ NPs and the NP mixture were aged in 600 ml SWW (SWW composition is described in detail in the Supplementary Information section; SI) in 1 L borosilicate glass jars (Flocculator 2000, Kemira Oyi, Finland) prior to use in exposure studies. The samples (n=3) were stirred rapidly (250 rpm) for 30 seconds to ensure homogeneous dispersion of the NP in the SWW, and were subsequently stirred slowly (50 rpm) for 4 h at 20 $\pm 2^{\circ}$ C. Samples were intermittently taken at time points 0 h, 1 h, 2 h and 4 h to characterize the

particle behavior in SWW over time. SWW-aged (4h) NP dispersions of Ag NPs, TiO₂ NPs 128 and the NP mixture used for exposure experiments were prepared by tenfold dilution in filtered 129 natural seawater. Pristine NP exposure dispersions were similarly prepared by diluting 130 respective stocks directly in filtered natural seawater. Exposure dispersions (n=3) of pristine 131 and aged Ag NPs (1 mg/L), TiO₂ NPs (10 mg/L) and the NP mixture (1 and 10 mg/L Ag and 132 TiO₂ NPs, respectively), were characterized with UV-vis and DLS at 0 h, 24 h, 48 h, and 72 h, 133 134 corresponding to the exposure duration (exposure dispersion renewals at t=72 h). At the end of the 4 h aging process, the exposure dispersions at the highest nominal exposure concentration 135 136 were quantified using ICP-MS to help interpret data from the effect studies on the development of the T. battagliai. Ag NP characterization and quantification methodology by STEM (in 137 synthetic wastewater) and sp-ICP-MS is described in the Supporting Information (SI). 138

Particle potential. The particle size and size distribution, 139 size and zeta agglomeration/aggregation, particle count and surface charge (zeta potential) during aging and 140 141 exposures were evaluated using a Zetasizer instrument (NanoZS Malvern Instruments, UK).

Particle concentration via UV-Vis. To determine NP concentrations with UV-vis (UV-mini 1240, Shimadzu, Japan), absorbance peaks for each particle type were determined by spectral scans (190-800 nm). Calibration curves for Ag NP and TiO₂ NP dilution series (MilliQ-water) were prepared using peak absorbance values. The samples were resuspended manually immediately prior to the analysis.

147 *Total and dissolved Ag and Ti concentration-ICP-MS*. Samples for total Ag and Ti content 148 determination were obtained as explained in the previous section. The presence of dissolved 149 species of Ag and Ti was obtained by centrifugal ultrafiltration of the samples through a 3 kDa 150 cut-off membrane at 5000 g for 1 h (Amicon Ultra-15 Centrifugal Filter Unit, Millipore). The 151 samples were acidified by ultrapure HNO₃:HF (5:1 v/v) and analyzed with ICP-MS (Agilent

152 8800, triple Quadrupole). For quality control, blanks and certified reference material (NCS DC
153 73349) were included in the analysis.

Sp-ICP-MS analysis. Sub-samples of Ag NPs during aging and exposures were stored frozen
at -20°C until analysis. Samples were vortexed for 30s and diluted with MilliQ water (to reach
an Ag concentration of approximately 5 ppt) prior to analysis. The analytical protocol and data
analysis (single particle RIKILT calculation tool²⁵) was performed as previously described^{9,25}
(detailed method description in SI).

159 Effects on *T. battagliai* nauplii development.

T. battagliai cultures were maintained in natural seawater (0.22 µm filtered, 35 ppt salinity) 160 obtained from the outer Oslofjord and were fed a mixed algal diet of Rhodomonas baltica and 161 *Isochrysis galbana*. The developmental assay was performed as previously described²⁶ with 162 slight modifications. Gravid females were separated from the cultures and nauplii <12 h post-163 164 release were individually placed in the well of a 24-well plate containing 2 ml of test solution (10 replicates per concentration). A concentration representing 10% of the aged stocks in 165 seawater was used as the highest exposure concentration (corresponding to nominal 166 concentrations of 1 mg/L Ag NPs, 10 mg/L TiO₂ NPs and 1 and 10 mg/L Ag and TiO₂ NPs, 167 respectively in the NP mixture). The effects of increasing concentrations of Ag NPs (0.06, 168 0.125, 0.25, 0.5, 1 mg/L), TiO₂ NPs (0.6, 1.25, 2.5, 5, 10 mg/L) and the corresponding NP 169 mixture were assessed over an exposure duration of 6 days. AgNO₃ (0.003, 0.006, 0.0125, 170 0.025, 0.05 mg/L) was used as an ionic control. The exposure dispersions were prepared from 171 172 the highest nominal exposure concentration and a concentrated cell suspension of R. baltica was added to reach $2 \cdot 10^5$ cells/ml (final concentration) prior to distribution to the exposure 173 wells. Exposure solutions containing R. baltica ($2 \cdot 10^5$ cells/ml final concentration) were 174 renewed on day 3. A SWW control (10% in seawater as per the other highest nominal exposure 175

concentrations) and a natural seawater control were included. The pH in controls and treatments were monitored at the beginning and end of the exposures and ranged from 8.0 to 8.2 while the dissolved oxygen levels ranged from 7.29 to 7.65 mg/L. The developmental stage of the copepods was evaluated daily and mortality or other abnormalities were recorded. The naupliar developmental rate (% nauplii reaching the copepodid stage at the end of the study compared to the controls) was calculated for each treatment.

182 NP uptake and detection in *T. battagliai*.

To study the uptake of particles, organisms exposed to 0.006 mg/L AgNO₃, 0.125 mg/L Ag 183 NPs and the mixture of Ag NPs (0.125 mg/L) and TiO₂ NPs (1.25 mg/L) were transferred to 184 clean natural seawater at the end of the exposure period (day 6) and allowed to depurate for 24 185 186 h. The concentrations were chosen based on the highest concentration that did not exert an effect. At the end of the depuration period, 5 individuals were pooled and transferred to 187 Eppendorf tubes. They were washed twice with a solution containing N-Acetyl-L-cysteine 188 (NAC, Sigma Aldrich, final concentration 0.5 mM in MilliQ water) to remove any loosely 189 bound ionic Ag or NPs on the surface of the organisms and the samples were stored at -20°C 190 191 until further processing. The digestion process was performed as previously described²⁵, but slightly modified and optimized for *T. battagliai* used in this study. Briefly, 0.2 ml of digestion 192 buffer (10 mM Tris buffer, 0.1% Triton X and 1 mM calcium acetate, pH 9) was added to each 193 194 tube containing 5 copepods and the samples were sonicated for 1 h in a sonication bath. The samples were then enzymatically digested by adding 1 µL of Proteinase K (Merck-Millipore, 195 final concentration 100 μ g/mL) to each tube and incubating the samples overnight at 37°C. The 196 197 samples were then sonicated for 15 min in a sonication bath and diluted in MilliQ water prior to sp-ICP-MS analysis (analysis was performed within 1 hour of sample dilution). In order to 198 assess the impact of the digestion process on the NP size, samples of the digestion buffer spiked 199

with Ag NPs according to the procedure described above were also analyzed with sp-ICP-MS and resulted in particles of 27 ± 1.4 nm. Although there was no visible debris after the enzymatic digestion, incomplete digestion of the exoskeleton cannot be excluded. Therefore, any NPs remaining (after the washing steps with NAC) associated with the exoskeleton were not taken into account in the sp-ICP-MS measurement.

205 Statistical Analysis.

Statistical analyses were performed with GraphPad Prism 6 (GraphPad Software, La Jolla, CA). Values are expressed as means ± standard deviation and the exposure experiments were repeated 2 to 3 times. Significant differences in concentrations between aged and pristine particles and between treatments and controls were analyzed with one-way ANOVA followed by Sidak's or Dunnet's multiple comparison test. Dose response curves and EC50 values were obtained with GraphPad Prism 6 (GraphPad Software, La Jolla, CA 92037) using a logistic four-parameter model.

213 RESULTS AND DISCUSSION

Stock dispersion characterization. DLS analysis showed that the pristine Ag NPs had an average hydrodynamic (z-ave) size of 66 ± 0.1 nm, while the TiO₂ NPs had a z-ave size of 358 ± 11 nm (Table 1), which is in accordance with our previous studies showing the presence of TiO₂ aggregates consisting of NP below 10 nm^{9,27}. According to sp-ICP-MS analysis, the average size of Ag NPs in the stock dispersion was 24 ± 3 nm (Figure S1). Due to the aggregate size of the TiO₂ particles in the stock dispersion, sp-ICP-MS was only performed for Ag NP stock dispersions.

Nanoparticle dispersion characterization during aging and exposure. An increase in the z ave was observed for both particles during aging in SWW, resulting in values of 381±44 nm

for the Ag NPs and 511 \pm 7.5 nm for the TiO₂ NPs after 4 h (Table 1). This may at least partly 223 be attributed to the presence of particles present in SWW, which had a z-average of 536 ± 34 224 nm (data not shown). A similar trend of increasing size was also previously reported for Ag 225 NPs during aging in crude wastewater²⁸. A previous study has shown that PVP-coated Ag NPs 226 were rapidly transformed to Ag(0) in influent wastewater (after 4 min of spiking), suggesting 227 that a modification/loss of the surface coating occurs²⁹. Despite the fact that this can vary 228 between different PVP-coated NPs, a potential modification of the surface coating in the 229 present study cannot be excluded and may have contributed to the aggregation observed in 230 231 SWW immediately after spiking.

232

Table 1. Characterization data for Ag NPs, TiO_2 NPs and the NP mixture at either: 0 h (start) and 4 h (end) of aging (in synthetic wastewater; SWW), or during exposure in natural seawater as aged or pristine at 0h (start of exposure) and 72h (exposure solution renewal). Values presented are the average \pm standard deviation of triplicates.

			SWW		Pristine NP in seawater		Aged NP in seawater	
		Stock	during	g aging	during	exposure	during	exposure
			0 h	4 h	0 h	72 h	0 h	72 h
	z average (nm)	66±0.1	391±30	381±44	75±2.5	90±11	268±36	316±37
Ag NPs	Size (number dist.; nm)	40±1.8	52±13	42±53	24±10	24±16	46±5.8	46±12
	Zeta potential (mV)	-28±1.7	-23±0.3	-17±8	-0.3±2.5	2.2±4.2	-8.6±1	-8.6±0.7
	z average (nm)	358±11	522±9.3	511±7.5	830±105	4234±321	548±41	821±65
TiO ₂ NPs	Size (number dist.; nm)	218±44	224±62	232±12	542±138	1500±460	234±12	232±39
	Zeta potential (mV)	-3.3±0.3	-20±0.8	-20±2.6	0.3±0.7	-1.1±0.9	-8±0.5	-9±2.1
Ag+TiO ₂	z average (nm)	-	470±3.4	454±3.4	641±17.1	1237±42	469±30	671±148
	Size (number dist.; nm)	-	120±59	147±117	3 69±185	675±54	141±38	132±63

Zeta potential (mV) - -19 ± 0.9 -15 ± 5.4 -1.5 ± 1 -2.2 ± 0.6 -6.2 ± 0.6 -6.1 ± 1

237

STEM analysis of Ag NPs and TiO₂ NPs upon co-exposure after 4h of aging in SWW 238 confirmed the presence of Ti aggregates of approximately 400 nm (Figure 1A) and Ag-rich 239 nanoparticles of approximately 25 nm and small agglomerates dispersed in clusters (Figure 240 1B). Moreover, areas with Ag NPs associated with the surface of TiO₂ NPs were also observed 241 (Figure 1C). Energy-dispersive X-ray spectroscopy (EDS) analysis showed an association of 242 Ag, Ti, Ca, and P indicating heteroaggregation of Ag NPs with TiO₂ NPs and components of 243 the wastewater. The elemental distribution suggests the presence of Ca and P at the surface 244 layers (Figure 1A, C), with the core regions showing heavier mass contrast. A slight increase 245 in Ag NP z-ave size was observed for pristine Ag NPs (from 75±2.5 to 90±11 nm) in seawater 246 after 72 h. The larger size of the aged Ag NPs remained in the exposure media (from 268±36 247 to 316±37 nm), showing the presence of SWW particles in the exposure media. Sp-ICP-MS 248 showed a size increase of SWW-aged Ag NPs in seawater over time from 28±0.4 nm to 249 31.4±0.4±1.6 nm (Figure S1). The differences in NP diameter measured with DLS and sp-ICP-250 MS are probably due to the higher concentrations needed for DLS that can lead to the aggregate 251 252 formation as well as the presence of SWW particles that can influence the signal^{9,30}. Therefore, sp-ICP-MS is a more suitable technique for metallic NP analysis in complex matrices at low 253 254 concentrations.





Figure 1. High Angle Annular Dark Field (HAADF) STEM images and elemental analysis of
Ag and TiO₂ particles (co-incubation) after 4h aging in wastewater. (A) Ti particles (in red)
associated with Ca (in blue), (B) Ag particles (in green) associated with Ca (in blue) and (C)
co-localization of Ti (in red) with Ag (in green) and Ca (in blue).

To determine the influence of SWW particles on the dispersion stability measurements, we included measurements of the pure SWW as background controls. The z-ave of SWW (inherent particles) in seawater increased from 395 ± 121 nm to 843 ± 499 nm, indicating agglomeration/aggregation of SWW particles occurs in seawater. After 72 h, the aggregate size and zeta potential of the pristine TiO₂ NPs in seawater were 4234 ± 321 nm and -1.1 ± 0.9 mV,

respectively, while they were 821 ± 65 nm and -9 ± 2.1 mV, respectively for the aged TiO₂ NPs. 266 The smaller size of aged TiO₂ NP and the size overlap with particles present in the pure SWW 267 when added to the exposure media again shows the influence of SWW particles and/or their 268 interactions with TiO₂ NPs. A similar behavior of smaller aggregate size formation and a 269 decrease in zeta potential after aging was observed for the mixture of Ag NP and TiO₂ NP in 270 seawater (671±148 nm). Our results clearly show the influence of SWW particles on size 271 272 measurements, which in the case of Ag NP leads to an increased z-ave, and for TiO₂ NP leads to a smaller z-ave in the exposure media. 273

274 Further dispersion stability measurements of aged and pristine NP in seawater performed with UV-vis spectroscopy showed that the effects on particle/aggregate stability are not caused by 275 SWW particles being present in the size measurements, but are also due to interactions with 276 the NPs with SWW components. The concentration of Ag and TiO₂ NPs in natural seawater in 277 the absence of aging decreased over time (Figure 2), indicating either changes in 278 279 agglomeration/aggregation state and sedimentation of the formed aggregates or increased dissolution (for Ag NP). A decrease in UV-vis absorbance of PVP-Ag NPs in seawater (30 ppt) 280 was attributed to NP dissolution according to a previous study²⁰. Aggregation of Ag NPs in 281 aqueous media of increasing ionic strength has been previously shown³¹. The conductivity (as 282 an indicator of ionic strength) of the seawater at 35 ppt used in the exposures was 46.7 mS/cm. 283 This leads to increased aggregation and settling, which could explain the decrease in 284 concentration and increase in hydrodynamic diameter of Ag and TiO₂ NP over time in 285 seawater. A higher decrease (60% decrease) in the Ag NP concentration was observed in the 286 287 presence of TiO₂ NPs in seawater compared to the single exposures suggesting that TiO₂ NPs leads to increased Ag NP co-precipitation (Figure S2). In accordance to this study, 288 heteroaggregation of Ag and TiO₂ NPs at higher TiO₂ NP concentration has been previously 289 reported³². Interestingly, an increase in dispersion stability was observed for both Ag NPs and 290

TiO₂ NPs when the particles were aged for 4 h in SWW, reaching 80% and 60% of the total concentration of Ag NPs and TiO₂ NPs, respectively after 72 h in exposure media (seawater) (Figure 2).



295

Figure 2. Impact of aging on (A) Ag NP (1 mg/L) and (B) TiO₂ NP (10 mg/L) dispersion stability in natural seawater over time. The concentration of pristine (freshy prepared, black bars) or aged (4 h in SWW, grey bars) NP in natural seawater was determined with UV-Vis at 0, 24, 48 and 72 h. The data is presented as mean \pm SD. Asterisks denote statistically significant differences (*p*<0.01).

301

Previous studies have demonstrated an increased dispersion stability of Ag NPs in moderately hard reconstituted water containing humic acids compared to media with higher chloride (Cl⁻) concentrations²¹. Moreover, it has been recently shown that TiO_2 NPs were more stable over time in natural water samples containing high molecular weight biopolymers³³ and in seawater in the presence of dissolved organic carbon despite the high salinity levels of 30 ppt³⁴. This could explain the increased dispersion stability observed in the case of the Ag NP aged in SWW that contains high levels of total organic carbon and proteins. In the current study natural 309 organic matter was not included in the synthetic wastewater which could lead to further 310 increase in stability depending on the natural organic matter quality and media ion 311 composition⁸.

Total, dissolved and particulate Ag. At the highest nominal exposure concentration of 1 312 mg/L, the Ag NP dissolution was observed to increase over time, reaching measured 313 concentrations of 441 µg/L and 378 µg/L, respectively for pristine and aged NP after 72h in 314 seawater (Figure 3A). This represents dissolution of 48±10 % and 49±2.8 % of the total Ag 315 added (1 mg/L) for pristine and aged, respectively. A low dissolution rate of approximately 4% 316 has previously been reported for Ag NPs (NM 300K, EU JRC Repository) in both bacterial 317 media and artificial wastewater³⁵. The lower dissolution reported is probably due to the shorter 318 experimental time of 1 h (compared to 72 h in the current study), as well as the higher 319 concentration used in the dissolution experiment (50 mg/L compared to 1 mg/L used in the 320 current study). The dissolution rate has previously been shown to be affected by both the 321 322 concentration and size of NPs, with a decrease in aggregate size and an increase in dissolution as the NP concentration decreases^{19–21}. Moreover, it has been previously shown that the Cl/Ag 323 ratio determines the dissolution rate with increasing Cl/Ag ratio leading to increased 324 dissolution and increased presence of $AgCl_x^{(x-1)}$ - soluble species compared to AgCl (s). In the 325 current study, the Cl/Ag ratio is 19400 and so the formation of soluble Ag-Cl complexes is 326 expected³⁶. Despite the observed increased stability during aging measured with UV-vis the 327 soluble Ag levels measured with ICP-MS were similar for both aged and pristine particles in 328 seawater. This discrepancy could result from interference in UV-vis absorbance due to 329 aggregation^{20,21}, sorption of constituents present in the media²⁰ and the presence of particles in 330 the SWW. It has been previously discussed that there is no ideal technique for the 331 characterization and detection of NPs in complex environmental matrices but rather a 332 combination of multiple analytical techniques is needed³⁰. 333



334

335 Figure 3. (A) Total and dissolved measured Ag concentrations of pristine and aged (in SWW) Ag NPs (nominally 1 mg/L) over time in seawater, either as single or combined exposures with 336 TiO₂ NPs (nominally 10 mg/L). (B) Measured NP concentration of aged Ag NPs (nominally 1 337 mg/L) in single exposures and in combination with TiO₂ NPs as measured over time using sp-338 ICP-MS. The total, dissolved Ag (centrifugal ultrafiltration using a 3kDa filter membrane-ICP-339 MS) and particulate form (sp-ICP-MS) were determined after 0 h and 72 h of exposure. The 340 values shown are measured concentrations. The error bars represent the standard deviation of 341 three replicates. Asterisks denote statistically significant differences (p < 0.05). 342

Relative to the Ag NP single exposures, a decrease in the dissolved Ag concentration was measured in samples containing the Ag NP and TiO_2 NP mixture in both pristine and aged exposures (Figure 3A). This indicates that the presence of TiO_2 either inhibits the dissolution process or that the dissolved species rapidly become associated with surface of the TiO_2 NPs. In accordance with the current findings, a previous study has shown that under illumination, TiO_2 NPs led to a 64.3% decrease in Ag⁺ release from Ag NPs compared to Ag NP only exposures and the reformation of Ag NPs on the surface of TiO_2 NPs²². Similarly, lower Ag⁺ levels were measured when Ag NPs and TiO_2 NPs were present as a mixture in lake water, where an increase in Ag⁺ sorption to TiO_2 particles was observed²³. It should be noted that the presence of Ag-containing solids that do not pass through the 3kDa membrane could be omitted from the analysis leading to an underestimation of the dissolved Ag species.

The behavior of Ag NPs during exposures of SWW-aged and pristine particles was also studied 355 with sp-ICP-MS, a technique that has low detection limits and can provide size and particle 356 concentration at environmentally relevant concentrations. In the exposure media, a decrease in 357 the Ag NP measured concentration from 237 µg/L to 96.9 µg/L (Figure 3B) was observed after 358 72h exposure (exposure media renewal), which is in accordance with the increased dissolution 359 with time. Similar results using sp-ICP-MS analysis have been previously reported, showing 360 increased Ag dissolution over time accompanied by a decrease in particle number that indicates 361 transformation to dissolved or complexed Ag or formation of smaller-sized Ag NPs³⁷. 362 Moreover, secondary particles were shown to form during Ag NP dissolution in complex 363 matrices such as wastewater effluent samples that contain high concentrations of amino acids³⁸. 364 The lower soluble Ag levels observed in the presence of TiO_2 in the current study, together 365 with the lower Ag NP concentration measured with sp-ICP-MS, indicate a reformation of Ag 366 particulate complexes with TiO₂ after dissolution or formation of smaller sized particles. Due 367 to the limit of detection of sp-ICP-MS, particles below 19 nm will be incorporated into the 368 dissolved fraction, therefore the presence of smaller-sized Ag complexes are not measured. 369

Effects of particles on *T. battagliai* naupliar development. A developmental rate of 0.28 d⁻¹ was determined in both natural seawater and SWW (10% SWW in natural seawater) controls and this value was set to represent 100%. An increased toxicity was observed upon exposure to the positive AgNO₃ control, resulting in an EC50 of 9.86 μ g/L (SI, Figure S2), which is in

the same range as in previous studies with the copepod Amphiascus tenuiremis³⁹ and confirms 374 the increased sensitivity of the naupliar developmental stage. Exposure to increasing 375 concentrations of Ag NPs led to delayed development at concentrations above 500 µg/L with 376 a complete cessation of molting and increased mortality (Figure 4A). The aging process led to 377 increased toxicity compared to the pristine Ag NPs with a 2-fold decrease in EC50 (316 μ g/L) 378 compared to pristine Ag NPs in natural seawater (EC50 = 640 μ g/L). At the highest exposure 379 380 concentration of 1 mg/L the measured dissolved Ag concentrations were at similar levels for both aged and pristine exposures (441 μ g/L and 378 μ g/L, respectively), suggesting that the 381 382 effects and toxicity cannot be predicted by the dissolved or total Ag concentrations but the exposure media composition and complexity should also be taken into account. Ag NP aging 383 in the complex SWW media containing high levels of organic matter could lead to increased 384 residence times in the water column and increased exposure. It has been previously shown that 385 Ag NPs spiked in crude wastewater resulted in lower toxicity to the bioluminescent bacteria 386 Pseudomonas putida compared to spiked final wastewater²⁸. The differences in toxicity 387 observed despite the similar Ag levels measured can be due to differences in interaction 388 between dissolved Ag and Ag ions/NPs attached to SWW particles during aging with different 389 tissues/external surfaces of the organism. 390





Figure 4. Effects of pristine vs aged NPs on the naupliar development of *T. battagliai*. The nauplii were exposed for 6 days to (A) Ag NPs (0-1 mg/L), (B) TiO₂ NPs (0-10 mg/L) and (C) a mixture of Ag and TiO₂ NPs at increasing concentrations. The exposure media were renewed after 72 h. The values shown are nominal exposure concentrations.

396 No effects on *T. battagliai* development rate or viability were observed upon exposure to

TiO₂ NPs despite the high exposure concentrations employed (0.64-10 mg/L) and irrespective

of the aging process (Figure 4B). Similar results have also been recently observed for

rainbow trout exposed to either wastewater-borne or water dispersed TiO_2 NPs where no

400 effects on mortality, behavior and growth were found¹⁶.

A similar pattern to that observed for the Ag NP only exposures was also observed when the 401 nauplii were exposed to the mixture of aged Ag and TiO₂ NPs (Figure 4C), with a dose-402 403 dependent effect on naupliar development and number of organisms reaching the copepodid stage leading to an EC50 of 270 µg/L. In the combined exposures, higher dissolved Ag levels 404 were measured for the aged Ag NPs (238.3 µg/L) compared to the pristine Ag NPs (98.57 405 μ g/L) (Figure 3A). Increased release of Ag ions in the presence of TiO₂ NPs was previously 406 described under sunlight irradiation with the simultaneous formation of Ag⁰ particles of smaller 407 size³². The increased toxicity may be due to one or a combination of increased dissolution, re-408 formation of smaller Ag particles and the presence of TiO₂ particles that combined lead to 409 enhanced effects through a different mode of action. In a previous study, TiO₂ led to enhanced 410 Ag NP toxicity to Tetrahymena pyriformis under illumination conditions, which was associated 411 with oxidative stress and a formation of Ag-TiO₂ particle complexes after Ag⁺ sorption to the 412 surface of TiO₂ particles²². Increased Ag toxicity to *Daphnia magna* has also been previously 413 observed in the presence of TiO₂ with higher Ag body burdens that were explained by ingestion 414 of Ag-sorbed TiO₂ NPs⁴⁰. In the current study, the observed differences in toxicity are 415

suggested to be due to the dynamic nature of the exposures, where organisms are being exposed 416 to (i) a mixture of both particles and dissolved species released over time and (ii) different 417 particulate complexes formed over time, which is in agreement with previous studies³⁹. It 418 should be noted that the NP behavior and subsequent exposure to T. battagliai may be altered 419 by the presence of the algae. NP agglomeration/interaction with released extracellular 420 polymeric substances and deposition can alter the actual exposure, while NP/ion interaction 421 422 with the algae surface during the exposures can also lead to decrease in the dissolved species measured³⁹. Moreover, the presence of food in the exposure system can lead to increased 423 424 bioavailability due to Ag ions/NPs with the food source and increased uptake⁴¹.

Ag NP uptake determination with sp-ICP-MS. The Ag NP concentration retained within T. 425 battagliai after 6 day exposure and a 24 h depuration period was evaluated with sp-ICP-MS in 426 organisms exposed to concentrations at which no developmental effects were observed; 125 427 μg/L pristine Ag NPs, 125 μg/L aged Ag NPs, mixture of 125 μg/L Ag and 1250 μg/L TiO₂ 428 429 NPs (aged), 6 µg/L AgNO₃ (nominal concentrations). Ag particles in the nanoscale were 430 detected in T. battagliai in all treatments, with an average size of 25 nm in treatments containing Ag NPs (alone or combined with TiO₂ NPs) and 40 nm in the AgNO₃ treatment 431 (Table 2). Among the particle exposures, the highest Ag concentration retained in the 432 organisms after 24 h depuration was found in the groups exposed to the pristine Ag NPs, where 433 3.45% of Ag NPs were retained (0.30 µg/copepod) relative to the Ag NP levels measured in 434 the exposure media (8.83 µg Ag NPs/L). Lower Ag NPs levels were measured in organisms 435 exposed to the aged Ag NPs, with a 2.4 times lower Ag NP percentage retention compared to 436 437 the pristine Ag NPs, reaching 0.08 µg Ag NPs/L (1.44% retention) and indicating a decreased bioavailability of the aged particles compared to the pristine counterparts. These results are in 438 accordance with previous studies showing that Ag bioavailability to the alga Chlamydomonas 439 440 reinhardtii is decreased in wastewaters compared to simplified media³⁷. Similarly, a limited

bioavailability of wastewater-borne Ag NPs to rainbow trout upon aqueous exposure compared
to the pristine counterparts has been reported presumably due to particle transformation⁴².
Despite the lower bioavailability, the complexed species and smaller Ag NP particles being
formed in wastewater media can still be taken up by the copepods.

445

446 **Table 2.** Overview of Ag NP content and size measurement with sp-ICP-MS in exposure

447 media and *T. battagliai* (measured concentrations) after 24 h of depuration, with and without

448 NP aging.

Exposure media					T. battagliai		
Fv	nocure	Size (nm)	Measured conc	Size (nm)	Measured conc (µg	% Ag NPs retained in organisms	
LA	posure	Size (IIIII)	(µg Ag NPs/L)	Size (IIII)	Ag NPs/copepod)		
Drigting	Ag	27.17	8.83±3.2	24.09	0.30±0.13	3.45	
Plistille	AgNO ₃	40.18	1.99±0.13	38.35	0.23±0.19	11.74	
Acad	Ag	25.53	5.83±1.48	25.11	0.08	1.44	
Aged	Ag +TiO ₂	24.53	5.23±1.03	19.08	0.15	2.82	

449

In the presence of TiO₂, a higher concentration of Ag NPs was retained in the organisms 450 compared to the Ag NP single exposure (~2 times higher percentage Ag NP retention). A higher 451 bioavailability for aqueous Ag compared to Ag in particulate form has been previously reported 452 for uptake studies using the algae C. vulgaris⁴³. Similarly, the highest Ag levels retained in the 453 organisms (11.7%) relative to the measured Ag levels in the exposure media were observed in 454 the AgNO₃ treatment indicating an increased bioavailability which is in accordance with 455 previous studies reporting higher Ag uptake in Daphnia magna in aqueous exposures of 456 AgNO₃ compared to Ag NPs⁴⁴. An average particle size of ~40 nm was measured in the 457 organisms exposed to AgNO₃ which suggests a particle formation within the organisms after 458

exposure to the ionic form. A biogenic formation of Ag NP forms from ionic Ag has been 459 previously detected in earthworms Lumbricus rubellus and Eisenia fetida tissues exposed to 460 $AgNO_3^{45,46}$ and it was suggested this represented a protection and detoxification mechanism. 461 The formation of granules containing metals, including Ag, with sulfur or phosphorus 462 precipitates within cell vacuoles has been shown to play an important role in metal 463 sequestration and detoxification process in invertebrates including crustaceans⁴⁷. The use of 464 465 sp-ICP-MS in combination with an enzymatic digestion method allowed the detection of Ag NPs in complex matrices such as whole organisms at low concentrations and helped towards 466 467 gaining an improved understanding of particle behavior in complex matrices and interpreting effects data. 468

Environmental Implications. The continuous NP release and enhanced toxicity that can be 469 observed in co-exposure with other NPs and the NP altered behavior in different environmental 470 media can lead to toxicity that cannot be explained or predicted based on total, soluble or 471 particulate concentrations. The nature of the NP behavior and association with organic 472 matter/biomolecules and the increased sensitivity observed for (epi)benthic organisms raise 473 concerns over the environmental impact of settling NP in natural systems. Importantly, the 474 complexity and environmental relevance of the matrix selected for use in exposure experiments 475 can have a significant impact on the dispersion stability, agglomeration/aggregation and 476 dissolution behavior of NPs and their subsequent effects on exposed organisms. The 477 concentrations used in the current study were higher than the ones currently found in the 478 environment, therefore future research efforts should focus on more environmentally relevant 479 480 aging conditions at environmentally relevant concentrations and in combination with other environmental stressors including co-contaminants and elucidate factors that lead to altered NP 481 bioavailability. 482

483 ASSOCIATED CONTENT

- 484 Supporting Information. Additional information is provided on synthetic wastewater
- 485 composition, NP characterization methodology (STEM/EDS, sp-ICP-MS), the size
- distribution of Ag NPs during aging and exposures obtained by sp-ICP-MS (Figure S1), UV-
- 487 vis measurements of pristine and aged Ag and TiO_2 NPs in combined exposures over time
- 488 (Figure S2), effect of AgNO₃ on *T. battagliai* naupliar development (Figure S3), Ag NP
- 489 characterization (size and concentration) during aging in SWW and exposure in SW over
- 490 time in single and combined exposures with TiO_2 NPs obtained by sp-ICP-MS (Table S1),
- 491 measured total Ti concentration of pristine and aged TiO_2 NPs in single and combined
- 492 exposures with Ag NPs at the start of exposures (Table S2).

493 AUTHOR INFORMATION

494 Corresponding Author

495 * Anastasia Georgantzopoulou. E-mail: <u>anastasia.georgantzopoulou@niva.no</u>. Tel:
496 +4798227741

497 Author Contributions

- The manuscript was written through contributions of all authors. All authors have givenapproval to the final version of the manuscript.
- 500

501 ACKNOWLEDGMENT

The authors would like to thank the Research Council of Norway (RCN) for the financial support, this study was performed within the RCN funded project NanoWASTE "Investigating the fate of nanomaterials in wastewater treatment plants; removal, release and subsequent impacts" (Grant Agreement number 238972/O70). This study has also been supported by FORURENS Nanoparticle Characterization in Environmental Media: Linking exposure to effects NANOCHARM (Researcher project – MILJØ2015). We thank Karina Pettersen
(Norwegian Institute for Water Research, Oslo, Norway) for her help with the maintenance of
the *T. battagliai* cultures and Valentina Zivanovic (NMBU, Ås, Norway) for the ICP-MS
analysis.

- 511 REFERENCES
- Weir, A.; Westerhoff, P.; Fabricius, L.; Hristovski, K.; Von Goetz, N. Titanium
 dioxide nanoparticles in food and personal care products. *Environ. Sci. Technol.* 2012,
 46 (4), 2242–2250.
- 515 (2) Voelker, D.; Schlich, K.; Hohndorf, L.; Koch, W.; Kuehnen, U.; Polleichtner, C.;

Kussatz, C.; Hund-Rinke, K. Approach on environmental risk assessment of nanosilver
released from textiles. *Environ. Res.* 2015, *140*, 661–672.

- (3) Polesel, F.; Farkas, J.; Kjos, M.; Almeida Carvalho, P.; Flores-Alsina, X.; Gernaey, K.
 V; Foss Hansen, S.; Plosz, B. G.; Booth, A. M. Occurrence, characterisation and fate
 of (nano)particulate Ti and Ag in two Norwegian wastewater treatment plants. *Water Res.* 2018, *141*, 19–31.
- 522 (4) Kiser, M. A.; Westerhoff, P.; Benn, T.; Wang, Y.; Pérez-Rivera, J.; Hristovski, K.

523 Titanium nanomaterial removal and release from wastewater treatment plants.

- 524 Environ. Sci. Technol. 2009, 43 (17), 6757–6763.
- 525 (5) Yang, Y.; Reed, R.; Schoepf, J.; Hristovski, K.; Herckes, P.; Westerhoff, P.
- Prospecting nanomaterials in aqueous environments by cloud-point extraction coupled
 with transmission electron microscopy. *Sci. Total Environ.* 2017, *584–585*, 515–522.
- 528 (6) Levard, C.; Hotze, E. M.; Lowry, G. V; Brown, G. E. Environmental transformations
- of silver nanoparticles: impact on stability and toxicity. *Environ. Sci. Technol.* **2012**,

- 530 *46* (13), 6900–6914.
- (7)Schultz, C.; Powell, K.; Crossley, A.; Jurkschat, K.; Kille, P.; Morgan, A. J.; Read, D.; 531 532 Tyne, W.; Lahive, E.; Svendsen, C.; Spurgeon, D. J. Analytical approaches to support current understanding of exposure, uptake and distributions of engineered 533 nanoparticles by aquatic and terrestrial organisms. Ecotoxicology 2015, 24, 239-261. 534 Metreveli, G.; Frombold, B.; Seitz, F.; Grün, A.; Philippe, A.; Rosenfeldt, R. R.; (8) 535 Bundschuh, M.; Schulz, R.; Manz, W.; Schaumann, G. E. Impact of chemical 536 composition of ecotoxicological test media on the stability and aggregation status of 537 silver nanoparticles. Environ. Sci. Nano 2016, 3, 418-433. 538 (9) 539 Georgantzopoulou, A.; Carvalho, P. A.; Vogelsang, C.; Ndungu, K.; Booth, A. M.; Thomas, K. V; Macken, A. Ecotoxicological effects of transformed silver and titanium 540 dioxide nanoparticles in the effluent from a lab-scale wastewater treatment system. 541 Environ. Sci. Technol. 2018, 52 (16), 9431–9441. 542 (10)Lin, S.; Taylor, A. a.; Ji, Z.; Chang, C. H.; Kinsinger, N. M.; Ueng, W.; Walker, S. L.; 543 544 Nel, A. E. Understanding the Transformation, Speciation, and Hazard Potential of Copper Particles in a Model Septic Tank System Using Zebrafish to Monitor the 545 Effluent. ACS Nano 2015, 9 (2), 2038–2048. 546 Muth-Köhne, E.; Sonnack, L.; Schlich, K.; Hischen, F.; Baumgartner, W.; Hund-(11)547 Rinke, K.; Schäfers, C.; Fenske, M. The toxicity of silver nanoparticles to zebrafish 548 549 embryos increases through sewage treatment processes. *Ecotoxicology* **2013**, *22* (8), 1264–1277. 550
- 551 (12) Kühr, S.; Schneider, S.; Meisterjahn, B.; Schlich, K.; Hund-Rinke, K.; Schlechtriem,
 552 C. Silver nanoparticles in sewage treatment plant effluents: chronic effects and

553		accumulation of silver in the freshwater amphipod Hyalella azteca. Environ. Sci. Eur.
554		2018 , <i>30</i> (1), 1–11.
555	(13)	Hartmann, S.; Louch, R.; Zeumer, R.; Steinhoff, B.; Mozhayeva, D.; Engelhard, C.;
556		Schönherr, H.; Schlechtriem, C.; Witte, K. Comparative multi-generation study on
557		long-term effects of pristine and wastewater-borne silver and titanium dioxide
558		nanoparticles on key lifecycle parameters in Daphnia magna. NanoImpact 2019, 14,
559		100163.
560	(14)	Kaegi, R.; Voegelin, A.; Sinnet, B.; Zuleeg, S.; Hagendorfer, H.; Burkhardt, M.;
561		Siegrist, H. Behavior of metallic silver nanoparticles in a pilot wastewater treatment
562		plant. Environ. Sci. Technol. 2011, 45 (9), 3902–3908.
563	(15)	Zhang, W.; Xiao, B.; Fang, T. Chemical transformation of silver nanoparticles in
564		aquatic environments: Mechanism, morphology and toxicity. Chemosphere 2018, 191
565		(7), 324–334.
566	(16)	Zeumer, R.; Galhano, V.; Monteiro, M. S.; Kuehr, S.; Knopf, B.; Meisterjahn, B.;
567		Soares, A. M. V. M.; Loureiro, S.; Lopes, I.; Schlechtriem, C. Chronic effects of
568		wastewater-borne silver and titanium dioxide nanoparticles on the rainbow trout
569		(Oncorhynchus mykiss). Sci. Total Environ. 2020, 723, 137974.
570	(17)	Kraas, M.; Schlich, K.; Knopf, B.; Wege, F.; Kägi, R.; Terytze, K.; Hund-Rinke, K.
571		Long-term effects of sulfidized silver nanoparticles in sewage sludge on soil
572		microflora. Environ. Toxicol. Chem. 2017, 36 (12), 3305–3313.
573	(18)	Kampe, S.; Kaegi, R.; Schlich, K.; Wasmuth, C.; Hollert, H.; Schlechtriem, C. Silver
574		nanoparticles in sewage sludge: Bioavailability of sulfidized silver to the terrestrial
575		isopod Porcellio scaber. Environ. Toxicol. Chem. 2018, 9999 (9999), 1-8.

576	(19)	Baalousha, M.; Sikder, M.; Prasad, A.; Lead, J.; Merrifield, R.; Chandler, G. T. The
577		concentration-dependent behaviour of nanoparticles. Env. chem 2016, 13, 1-3.
578	(20)	Sikder, M.; Lead, J. R.; Chandler, G. T.; Baalousha, M. A rapid approach for
579		measuring silver nanoparticle concentration and dissolution in seawater by UV-Vis.
580		Sci. Total Environ. 2018, 618, 597–607.
581	(21)	Zook, J. M.; Long, S. E.; Cleveland, D.; Geronimo, C. L. A.; MacCuspie, R. I.
582		Measuring silver nanoparticle dissolution in complex biological and environmental
583		matrices using UV-visible absorbance. Anal. Bioanal. Chem. 2011, 401 (6), 1993-
584		2002.
585	(22)	Zou, X.; Shi, J.; Zhang, H. Coexistence of silver and titanium dioxide nanoparticles:
586		Enhancing or reducing environmental risks? Aquat. Toxicol. 2014, 154, 168–175.
587	(23)	Wilke, C. M.; Tong, T.; Gaillard, JF.; Gray, K. A. Attenuation of Microbial Stress
588		Due to Nano-Ag and Nano-TiO ₂ Interactions under Dark Conditions. <i>Environ. Sci.</i>
589		Technol. 2016, 50, 11302–11310.
590	(24)	Jensen, K.; Booth, A.; Kembouche, Y.; Boraschi, D. NANoREG Deliverable 2.06.
591		Validated protocols for test item preparation for key in vitro and ecotoxicity studies.
592		<i>NANoREG</i> . 2016, p 77.
593	(25)	Peters, R.; Herrera-Rivera, Z.; Undas, A.; Van Der Lee, M.; Marvin, H.; Bouwmeester,
594		H.; Weigel, S. Single particle ICP-MS combined with a data evaluation tool as a
595		routine technique for the analysis of nanoparticles in complex matrices. J. Anal. At.
596		Spectrom. 2015, 30, 1274–1285.
597	(26)	Macken, A.; Lillicrap, A.; Langford, K. Benzoylurea pesticides used as veterinary

598		medicines in aquaculture: Risks and developmental effects on nontarget crustaceans.
599		Environ. Toxicol. Chem. 2015, 34 (7), 1533–1542.
600	(27)	Joint Research Centre of the European Commission. Titanium Dioxide, NM-100, NM-
601		101, NM-102, NM-103, NM-104, NM-105: Characterisation and Physico- Chemical
602		Properties; 2014.
603	(28)	Mallevre, F.; Alba, C.; Milne, C.; Gillespie, S.; Fernandes, T.; Aspray, T. Toxicity
604		Testing of Pristine and Aged Silver Nanoparticles in Real Wastewaters Using
605		Bioluminescent Pseudomonas putida. Nanomaterials 2016, 6 (3), 49.
606	(29)	Doolette, C. L.; Mclaughlin, M. J.; Kirby, J. K.; Batstone, D. J.; Harris, H. H.; Ge, H.;
607		Conrelis, G. Transformation of PVP coated silver nanoparticles in a simulated
608		wastewater treatment process and the effect on microbial communities. Chem. Cent. J.
609		2013 , 7 (46), 1–18.
610	(30)	Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J.
610 611	(30)	Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J. R.; Leppard, G. G.; Wilkinson, K. J. Characterizing Manufactured Nanoparticles in the
610 611 612	(30)	Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J.R.; Leppard, G. G.; Wilkinson, K. J. Characterizing Manufactured Nanoparticles in theEnvironment: Multimethod Determination of Particle Sizes. <i>Environ. Sci. Technol.</i>
610 611 612 613	(30)	 Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J. R.; Leppard, G. G.; Wilkinson, K. J. Characterizing Manufactured Nanoparticles in the Environment: Multimethod Determination of Particle Sizes. <i>Environ. Sci. Technol.</i> 2009, <i>43</i>, 7277–7284.
610 611 612 613 614	(30)	 Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J. R.; Leppard, G. G.; Wilkinson, K. J. Characterizing Manufactured Nanoparticles in the Environment: Multimethod Determination of Particle Sizes. <i>Environ. Sci. Technol.</i> 2009, <i>43</i>, 7277–7284. Badawy, A. M. E. L.; Luxton, T. P.; Silva, R. G.; Scheckel, K. G.; Suidan, M. T.;
610 611 612 613 614 615	(30)	 Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J. R.; Leppard, G. G.; Wilkinson, K. J. Characterizing Manufactured Nanoparticles in the Environment: Multimethod Determination of Particle Sizes. <i>Environ. Sci. Technol.</i> 2009, <i>43</i>, 7277–7284. Badawy, A. M. E. L.; Luxton, T. P.; Silva, R. G.; Scheckel, K. G.; Suidan, M. T.; Tolaymat, T. M. Impact of Environmental Conditions pH , Ionic Strength, and
 610 611 612 613 614 615 616 	(30)	 Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J. R.; Leppard, G. G.; Wilkinson, K. J. Characterizing Manufactured Nanoparticles in the Environment: Multimethod Determination of Particle Sizes. <i>Environ. Sci. Technol.</i> 2009, <i>43</i>, 7277–7284. Badawy, A. M. E. L.; Luxton, T. P.; Silva, R. G.; Scheckel, K. G.; Suidan, M. T.; Tolaymat, T. M. Impact of Environmental Conditions pH , Ionic Strength, and Electrolyte Type) on the Surface Charge and Aggregation of Silver Nanoparticles
 610 611 612 613 614 615 616 617 	(30)	 Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J. R.; Leppard, G. G.; Wilkinson, K. J. Characterizing Manufactured Nanoparticles in the Environment: Multimethod Determination of Particle Sizes. <i>Environ. Sci. Technol.</i> 2009, <i>43</i>, 7277–7284. Badawy, A. M. E. L.; Luxton, T. P.; Silva, R. G.; Scheckel, K. G.; Suidan, M. T.; Tolaymat, T. M. Impact of Environmental Conditions pH , Ionic Strength, and Electrolyte Type) on the Surface Charge and Aggregation of Silver Nanoparticles Suspensions. <i>Environ. Sci. Technol.</i> 2010, <i>44</i> (4), 1260–1266.
 610 611 612 613 614 615 616 617 618 	(30) (31) (32)	 Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J. R.; Leppard, G. G.; Wilkinson, K. J. Characterizing Manufactured Nanoparticles in the Environment: Multimethod Determination of Particle Sizes. <i>Environ. Sci. Technol.</i> 2009, <i>43</i>, 7277–7284. Badawy, A. M. E. L.; Luxton, T. P.; Silva, R. G.; Scheckel, K. G.; Suidan, M. T.; Tolaymat, T. M. Impact of Environmental Conditions pH , Ionic Strength, and Electrolyte Type) on the Surface Charge and Aggregation of Silver Nanoparticles Suspensions. <i>Environ. Sci. Technol.</i> 2010, <i>44</i> (4), 1260–1266. Zhang, Y.; Qiang, L.; Yuan, Y.; Wu, W.; Sun, B.; Zhu, L. Impacts of titanium dioxide
 610 611 612 613 614 615 616 617 618 619 	(30) (31) (32)	 Domingos, R. F.; Baalousha, M. A.; Ju-Nam, Y.; Reid, M. M.; Tufenkji, N.; Lead, J. R.; Leppard, G. G.; Wilkinson, K. J. Characterizing Manufactured Nanoparticles in the Environment: Multimethod Determination of Particle Sizes. <i>Environ. Sci. Technol.</i> 2009, <i>43</i>, 7277–7284. Badawy, A. M. E. L.; Luxton, T. P.; Silva, R. G.; Scheckel, K. G.; Suidan, M. T.; Tolaymat, T. M. Impact of Environmental Conditions pH , Ionic Strength, and Electrolyte Type) on the Surface Charge and Aggregation of Silver Nanoparticles Suspensions. <i>Environ. Sci. Technol.</i> 2010, <i>44</i> (4), 1260–1266. Zhang, Y.; Qiang, L.; Yuan, Y.; Wu, W.; Sun, B.; Zhu, L. Impacts of titanium dioxide nanoparticles on transformation of silver nanoparticles in aquatic environments.

621	(33)	Topuz, E.; Traber, J.; Sigg, L.; Talinli, I. Agglomeration of Ag and TiO ₂ nanoparticles
622		in surface and wastewater : Role of calcium ions and of organic carbon fractions.
623		Environ. Pollut. 2015, 204, 313–323.
624	(34)	Wang, H.; Burgess, R.; Cantwell, M.; Portis, L.; Perron, M.; Wu, F.; Ho, K. T.
625		Stability and aggregation of silver and titanium dioxide nanoparticles in seawater: role
626		of salinity and dissolved organic carbon. Environ. Toxicol. Chem. 2014, 33 (5), 1023-
627		1029.
628	(35)	Mallevre, F.; Fernandes, T. F.; Aspray, T. J. Silver, zinc oxide and titanium dioxide
629		nanoparticle ecotoxicity to bioluminescent Pseudomonas putida in laboratory medium
630		and artificial wastewater. Environ. Pollut. 2014, 195, 218-225.
631	(36)	Levard, C.; Mitra, S.; Yang, T.; Jew, A. D.; Badireddy, A. R.; Lowry, G. V.; Brown,
632		G. E. Effect of Chloride on the Dissolution Rate of Silver Nanoparticles and Toxicity
633		to E. coli. Environ. Sci. Technol. 2013, 47, 5738–5745.
634	(37)	Azimzada, A.; Tufenkji, N.; Wilkinson, K. J. Transformations of silver nanoparticles
635		in wastewater effluents: links to Ag bioavailability. Environ. Sci. Nano 2017, 4, 1339-
636		1349.
637	(38)	Azodi, M.; Sultan, Y.; Ghoshal, S. Dissolution Behavior of Silver Nanoparticles and
638		Formation of Secondary Silver Nanoparticles in Municipal Wastewater by Single-
639		Particle ICP-MS. Environ. Sci. Technol. 2016, 50 (24), 13318–13327.
640	(39)	Sikder, M.; Eudy, E.; Chandler, G. T.; Baalousha, M. Comparative study of dissolved
641		and nanoparticulate Ag effects on the life cycle of an estuarine meiobenthic copepod,
642		Amphiascus tenuiremis. Nanotoxicology 2018, 12, 1–15.

643	(40)	Rosenfeldt, R. R.; Seitz, F.; Schulz, R.; Bundschuh, M. Heavy Metal Uptake and
644		Toxicity in the Presence of Titanium Dioxide Nanoparticles: A Factorial Approach
645		Using Daphnia magna. Environ. Sci. Technol. 2014, 48, 6965–6972.
646	(41)	Kleiven, M.; Rossbach, L. M.; Gallego-urrea, J. A.; Brede, D. A.; Oughton, D.;
647		Coutris, C. Characterizing the behavior, uptake, and toxicity of NM300K silver
648		nanoparticles in Caenorhabditis elegans. Environ. Toxicol. Chem. 2018, 37 (7), 1799-
649		1810.
650	(42)	Zeumer, R.; Hermsen, L.; Kaegi, R.; Kühr, S.; Knopf, B.; Schlechtriem, C.
651		Bioavailability of silver from wastewater and planktonic food borne silver
652		nanoparticles in the rainbow trout Oncorhynchus mykiss. Sci. Total Environ. 2020,
653		706, 135695.
654	(43)	Kalman, J.; Paul, K. B.; Khan, F. R.; Stone, V.; Fernandes, T. F. Characterisation of
655		bioaccumulation dynamics of three differently coated silver nanoparticles and aqueous
656		silver in a simple freshwater food chain. Env. Chem. 2015, 12 (6), 662-672.
657	(44)	Ribeiro, F.; Gestel, C. A. M. Van; Pavlaki, M. D.; Azevedo, S.; Soares, A. M. V. M.;
658		Loureiro, S. Bioaccumulation of silver in Daphnia magna: Waterborne and dietary
659		exposure to nanoparticles and dissolved silver. Sci. Total Environ. 2017, 574, 1633-
660		1639.
661	(45)	Baccaro, M.; Undas, A. K.; De Vriendt, J.; Van Den Berg, J. H. J.; Peters, R. J. B.;
662		Van Den Brink, N. W. Ageing, dissolution and biogenic formation of nanoparticles:
663		How do these factors affect the uptake kinetics of silver nanoparticles in earthworms?
664		Environ. Sci. Nano 2018, 5 (5), 1107–1116.
665	(46)	Makama, S.; Peters, R.; Undas, A.; Van den Brink, N. W. A novel method for the

666		quantification, characterisation and speciation of silver nanoparticles in earthworms
667		exposed in soil. Env. Chem. 2015, 12, 643-651.
668	(47)	Ahearn, G. A.; Mandal, P. K.; Mandal, A. Mechanisms of heavy-metal sequestration
669		and detoxification in crustaceans: A review. J. Comp. Physiol. B 2004, 174 (6), 439-
670		452.

